

CHITOSAN-INDUCED CELL DEATH IN SYCAMORE CULTURED CELLS

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Programmed cell death (PCD) plays a pivotal role in many developmental plant processes and in defense mechanisms against biotic and abiotic stresses. At least three different forms of PCD have been reported in plants: a “nuclear” (apoptotic-like) form, a “chloroplastic” form and a “vacuolar” form¹. In plant cell cultures different stress conditions induce cell death that only in a fraction of the dead cells presents the typical morphological hallmarks of apoptosis, i.e. cell shrinkage, chromatin condensation, DNA fragmentation^{2,3}. Recently, our attention has been focused on chitosan (CHT). CHT is a natural, non-toxic and inexpensive compound obtained by partial alkaline deacetylation of chitin, the main component of the exoskeleton of crustaceans and other arthropods as well as of the cell wall of many fungi⁴. Although the exact mode of action of CHT is still unknown, in agriculture it has been shown to be a versatile compound that controls numerous pre and postharvest diseases of various horticultural commodities⁵. In sycamore (*Acer pseudoplatanus* L.) cultured cells CHT rapidly induces a set of defense/stress responses that include accumulation of dead cells and of cells with fragmented DNA accompanied by release of cytochrome *c* from the mitochondrion⁶. In this work we further investigated the cell death process induced by CHT. In particular, we tested the capability of CHT to induce oxidative stress (superoxide anion and malondialdehyde production) and to increase the activity of caspase3-like proteases.

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